

# Cytochrome *cbb*<sub>3</sub> of *Thioalkalivibrio* is a Na<sup>+</sup>-pumping cytochrome oxidase

Maria S. Muntyan<sup>a,1</sup>, Dmitry A. Cherepanov<sup>a</sup>, Anssi M. Malinen<sup>b</sup>, Dmitry A. Bloch<sup>a,c,2</sup>, Dmitry Y. Sorokin<sup>d,e</sup>, Inna I. Severina<sup>a,3</sup>, Tatiana V. Ivashina<sup>f</sup>, Reijo Lahti<sup>b</sup>, Gerard Muyzer<sup>g</sup>, and Vladimir P. Skulachev<sup>a,1</sup>

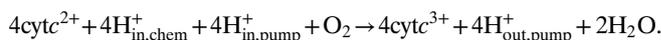
<sup>a</sup>Belozersky Institute of Physico-Chemical Biology, Lomonosov Moscow State University, Moscow 119991, Russia; <sup>b</sup>Department of Biochemistry, University of Turku, 20014 Turku, Finland; <sup>c</sup>Institute of Biotechnology, University of Helsinki, 00014 Helsinki, Finland; <sup>d</sup>Winogradsky Institute of Microbiology, Russian Academy of Sciences, Moscow 117312, Russia; <sup>e</sup>Department of Biotechnology, Delft University of Technology, 2628 BC Delft, The Netherlands; <sup>f</sup>Skryabin Institute of Biochemistry and Physiology of Microorganisms, Russian Academy of Sciences, Pushchino 142290, Russia; and <sup>g</sup>Microbial Systems Ecology, Department of Aquatic Microbiology, Institute for Biodiversity and Ecosystem Dynamics, University of Amsterdam, 1098 XH Amsterdam, The Netherlands

Edited by Harry B. Gray, California Institute of Technology, Pasadena, CA, and approved May 15, 2015 (received for review September 4, 2014)

**Cytochrome c oxidases (Coxs) are the basic energy transducers in the respiratory chain of the majority of aerobic organisms. Coxs studied to date are redox-driven proton-pumping enzymes belonging to one of three subfamilies: A-, B-, and C-type oxidases. The C-type oxidases (*cbb*<sub>3</sub> cytochromes), which are widespread among pathogenic bacteria, are the least understood. In particular, the proton-pumping machinery of these Coxs has not yet been elucidated despite the availability of X-ray structure information. Here, we report the discovery of the first (to our knowledge) sodium-pumping Cox (Scox), a *cbb*<sub>3</sub> cytochrome from the extremely alkaliphilic bacterium *Thioalkalivibrio versutus*. This finding offers clues to the previously unknown structure of the ion-pumping channel in the C-type Coxs and provides insight into the functional properties of this enzyme.**

cytochrome c oxidase | sodium pumping | *cbb*<sub>3</sub>-type oxidase | alkaliphily

The known terminal oxidases according to the structure of their active centers and their phylogenetic relations are subdivided into two superfamilies (1). One is composed of numerous representatives containing a heme-copper binuclear active center (BNC). Oxidases belonging to the other superfamily



In A-type Coxs, two H<sup>+</sup> pathways in the main subunit were identified, the so-called D channel, conducting all pumped and part of chemical H<sup>+</sup>, and the K channel, conducting most of chemical H<sup>+</sup> (9). In C-type Coxs, only a K-channel analog was found (10). The described catalytic events are accomplished through generation of a transmembrane difference in H<sup>+</sup> potentials ( $\Delta\bar{\mu}_{\text{H}^{+}}$ ), which is used as a convertible membrane-linked biological currency. Microorganisms living in an alkaline environment maintain a nearly neutral cytoplasmic pH (11). This presents a problem for alkaliphiles because it gives rise to an inverted pH gradient that decreases the  $\Delta\bar{\mu}_{\text{H}}$



## Significance

The majority of aerobic living organisms use oxygen for respiration. The key enzyme, which directly reduces oxygen to water during respiration, is the terminal cytochrome c oxidase. It generates a large portion of the utilizable energy provided by the respiratory chain. Accumulation of biologically available energy by means of cytochrome c oxidases is believed to be due to the proton-motive force across the mitochondrial or bacterial membrane. Details of this energy conversion are still unclear. Here we report the discovery of a sodium-pumping cytochrome c oxidase that converts energy of respiration into sodium-motive force. This finding provides clues to understanding the mechanism of cytochrome c oxidase that is not available when applying knowledge of the proton-pumping versions of the enzyme.

Author contributions: M.S.M., D.A.B., R.L., and V.P.S. designed research; M.S.M., A.M.M., D.A.B., D.Y.S., I.I.S., T.V.I., and G.M. performed research; M.S.M. computed molecular modeling; D.A.C. performed molecular dynamic computation and molecular modeling; M.S.M., D.A.C., A.M.M., D.A.B., D.Y.S., I.I.S., T.V.I., R.L., G.M., and V.P.S. analyzed data; and M.S.M., D.A.C., D.A.B., and V.P.S. wrote the paper.

The authors declare no conflict of interest.

This article is a PNAS Direct Submission.

Data deposition: The nucleotide sequences of *Thioalkalivibrio cbb*<sub>3</sub> oxidase have been deposited in the EMBL database (accession no. [HE575403.1](https://www.ebi.ac.uk/EMBL/nuclseq/HE575403.1)).

<sup>1</sup>To whom correspondence may be addressed. Email: [muntyan@genebee.msu.ru](mailto:muntyan@genebee.msu.ru) or [skulach@genebee.msu.ru](mailto:skulach@genebee.msu.ru).

<sup>2</sup>Deceased March 13, 2014.

<sup>3</sup>Deceased November 9, 2012.

This article contains supporting information online at [www.pnas.org/lookup/suppl/doi:10.1073/pnas.1417071112/-DCSupplemental](http://www.pnas.org/lookup/suppl/doi:10.1073/pnas.1417071112/-DCSupplemental).



- Gribaldo S, Talla E, Brochier-Armanet C (2009) Evolution of the haem copper oxidases superfamily: a rooting tale. *Trends Biochem Sci* 34(8):375–381.
- Rauhamäki V, Bloch DA, Verkhovsky MI, Wikström M (2009) Active site of cytochrome *cbb*<sub>3</sub>. *J Biol Chem* 284(17):11301–11308.
- Rauhamäki V, Baumann M, Soliymani R, Puustinen A, Wikström M (2006) Identification of a histidine-tyrosine cross-link in the active site of the *cbb*<sub>3</sub>-type cytochrome *c* oxidase from *Rhodobacter sphaeroides*. *Proc Natl Acad Sci USA* 103(44):16135–16140.
- Oztürk M, Watmough NJ (2011) Mutagenesis of tyrosine residues within helix VII in subunit I of the cytochrome *cbb*<sub>3</sub> oxidase from *Rhodobacter capsulatus*. *Mol Biol Rep* 38(5):3319–3326.
- Kaila VRI, Johansson MP, Sundholm D, Laakkonen L, Wikström M (2009) The chemistry of the Cu<sub>B</sub> site in cytochrome *c* oxidase and the importance of its unique His-Tyr bond. *Biochim Biophys Acta* 1787(4):221–233.
- Verkhovsky MI, Jasaitis A, Verkhovskaya ML, Morgan JE, Wikström M (1999) Proton translocation by cytochrome *c* oxidase. *Nature* 400(6743):480–483.
- Rauhamäki V, Bloch DA, Wikström M (2012) Mechanistic stoichiometry of proton translocation by cytochrome *cbb*<sub>3</sub>. *Proc Natl Acad Sci USA* 109(19):7286–7291.
- Rauhamäki V, Wikström M (2014) The causes of reduced proton-pumping efficiency in type B and C respiratory heme-copper oxidases, and in some mutated variants of type A. *Biochim Biophys Acta* 1837(7):999–1003.
- Konstantinov AA, Siletsky S, Mitchell D, Kaulen A, Gennis RB (1997) The roles of the two proton input channels in cytochrome *c* oxidase from *Rhodobacter sphaeroides* probed by the effects of site-directed mutations on time-resolved electrogenic intraprotein proton transfer. *Proc Natl Acad Sci USA* 94(17):9085–9090.
- Hemp J, et al. (2007) Comparative genomics and site-directed mutagenesis support the existence of only one input channel for protons in the C-family (*cbb*<sub>3</sub> oxidase) of heme-copper oxygen reductases. *Biochemistry* 46(35):9963–9972.
- Slonczewski JL, Fujisawa M, Dopson M, Krulwich TA (2009) Cytoplasmic pH measurement and homeostasis in bacteria and archaea. *Adv Microb Physiol* 55:1–79, 317.
- Skulachev VP (1984) Sodium bioenergetics. *Trends Biochem Sci* 9(11):483–485.
- Buschmann S, et al. (2010) The structure of *cbb*<sub>3</sub> cytochrome oxidase provides insights into proton pumping. *Science* 329(5989):327–330.
- Humphrey W, Dalke A, Schulten K (1996) VMD: Visual molecular dynamics. *J Mol Graph* 14(1):33–38, 27–28.
- Ekici S, Pawlik G, Lohmeyer E, Koch H-G, Daldal F (2012) Biogenesis of *cbb*<sub>3</sub>-type cytochrome *c* oxidase in *Rhodobacter capsulatus*. *Biochim Biophys Acta* 1817(6):898–910.
- Linding R, et al. (2003) Protein disorder prediction: Implications for structural proteomics. *Structure* 11(11):1453–1459.
- DeLano WL (2002) PyMOL Molecular Viewer. Available at [www.pymol.org](http://www.pymol.org).
- Kaila VRI, Verkhovsky MI, Wikström M (2010) Proton-coupled electron transfer in cytochrome oxidase. *Chem Rev* 110(12):7062–7081.
- Sharma V, Wikström M, Kaila VRI (2012) Dynamic water networks in cytochrome *cbb*<sub>3</sub> oxidase. *Biochim Biophys Acta* 1817(5):726–734.
- Ahn YO, et al. (2014) Conformational coupling between the active site and residues within the K<sup>C</sup>-channel of the *Vibrio cholerae* *cbb*<sub>3</sub>-type (C-family) oxygen reductase. *Proc Natl Acad Sci USA* 111(42):E4419–E4428.
- Ducluzeau A-L, Ouchane S, Nitschke W (2008) The *cbb*<sub>3</sub> oxidases are an ancient innovation of the domain bacteria. *Mol Biol Evol* 25(6):1158–1166.
- Schopf JW (2014) Geological evidence of oxygenic photosynthesis and the biotic response to the 2400–2200 Ma “Great oxidation event”. *Biochemistry (Mosc)* 79(3):165–177.
- Sousa FL, Alves RJ, Pereira-Leal JB, Teixeira M, Pereira MM (2011) A bioinformatics classifier and database for heme-copper oxygen reductases. *PLoS ONE* 6(4):e19117.
- Saraste M, Castresana J (1994) Cytochrome oxidase evolved by tinkering with denitrification enzymes. *FEBS Lett* 341(1):1–4.
- Mulkidjanian AY, Galperin MY, Makarova KS, Wolf YI, Koonin EV (2008) Evolutionary primacy of sodium bioenergetics. *Biol Direct* 3:13.

Electrical membrane potential generation in right-side-out membrane vesicles was monitored by the safranin method (49) or by tetraphenylphosphonium-selective electrode (50) at 25 °C.

H<sup>+</sup> release in intact cells and membrane vesicles in O<sub>2</sub>-pulse experiments was assessed by a standard method (51) in 1 mL of anoxic incubation mixture. Respiration of samples was initiated by addition of water (5–20 μL) saturated with air O<sub>2</sub> at 25 °C. The evoked changes in pH in the incubation mixture were estimated by titration with argon-saturated 0.5 mM H<sub>2</sub>SO<sub>4</sub>. Respiratory activity was assessed using a Clark-type electrode at 25 °C.

**ACKNOWLEDGMENTS.** We thank S. Tölkö, H. Luoto, S. Klishin, and D. Morozov for technical support, and M. Verkhovsky and V. Rauhamäki for providing *P. denitrificans* strains. This work was supported by the European Research Council Advanced Grant PARASOL 322551 (to G.M.); the Russian Foundation for Basic Research Grants 14-04-01577 (to M.S.M.), 05-04-49504 (to M.S.M. and D.A.B.), and 13-04-40405 (to D.Y.S.); and the Russian Scientific Fund Grant 14-50-00029 (to V.P.S.).

